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Temporal dynamics of planktonic microeukaryotes and bacteria in shallow soda lakes of the Carpathian Basin

-PhD thesis-

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#### 1. Introduction

Soda lakes are a special type of athalassic saline lakes, characterized by carbonate (CO<sub>3</sub><sup>2-</sup>), bicarbonate (HCO<sub>3</sub><sup>-</sup>), and sodium (Na<sup>+</sup>) as dominant ions in their water, and they have a stable alkaline pH (Boros et al., 2014; Felföldi, 2020). They can be found worldwide but they are less frequent than other naturalsaline lakes (Sorokin et al., 2014). In Europe, they predominantly occur as soda pans in the Carpathian Basin (Austria, Hungary, and Serbia) (Boros et al., 2014). These locally confined unique ecosystems serve as important feeding and breeding sites for waterbirds (Szabó et al., 2022) and as refuges for rare and endangered species (Horváth et al., 2013). Due to their uniqueness and their special biota, they are under legal protection, and most of them are located in national parks (Boros et al., 2017; 2013). Human activities, such as poor water management and various water drainages, have led to the disappearance of many soda lakes in the Carpathian Basin, which resulted in species loss as well (Boros et al., 2013; Felföldi, 2020; Horváth et al., 2019). Furthermore, Boros et al., (2020) implicated that the intensifying effects of climate change, such as prolonged desiccations due to decreasing precipitation and increasing temperature are threatening these vulnerable ecosystems.

There are previous studies that investigated the seasonality of the planktonic communities of soda lakes of the Carpathian Basin, for example in spring of 2013-2014, Szabó et al., (2020) observed sudden seasonal changes in the prokaryotic community composition. Pálffy et al. (2014) and Somogyi et al. (2009) found that in the summer, picocyanobacteria can dominate the water, while below 15 °C picoeukaryotic algae can cause mass production. Sometimes dual blooms of green algae and purple bacteria was also observed in these shallow soda pans (Korponai et al., 2019). But still we have very little information about the seasonality of the planktonic microbial communities and to what degree is seasonality determined by environmental variables and what extent desiccation influences the community composition.

Our goal was to gain knowledge about the planktonic microbial diversity along environmental gradients, and to get a comprehensive picture of the seasonality and community structure of micoruekaryotes and prokaryotes of soda pans.

#### 2. Main objectives

To date, a comprehensive study focusing on the planktonic microeukaryotes and prokaryotes of shallow soda lakes has not been carried out, which examines both the taxonomic composition, core microbiome (taxa shared among all sites), and diversity along environmental gradients as well as delving into their seasonal dynamics and ecological interactions. However, soda pans, since their community structure quickly responds to rapidly changing environmental parameters, can serve as models for understanding the processes of more complex aquatic ecosystems. Therefore, we studied the planktonic microbial communities through time (three seasons: spring, summer, and autumn by sampling biweekly) and space: (twenty-six soda pans by sampling in two consecutive spring seasons).

Accordingly, the main questions of my doctoral thesis were the following in the two projects:

A. Seasonal changes of the microeukaryotic and bacterial communities

I.) How similar are the seasonal changes of planktonic microeukaryotic and prokaryotic communities in nearby soda pans?

II.) How core and non-core microbial taxa contribute to the adaptation of the microbial communities, and how does this contribution vary between microeukaryotic and bacterial communities?

B. Temporal dynamics of microbial diversity along environmental gradients

III.) Is the identity and strength of the main environmental drivers change between subsequent years?

# 3. Materials and methods

## 3.1. Seasonal changes of the microeukaryotic and bacterial communities

- Biweekly sampling of five soda pans of Kiskunság from April 12 to November 14 in 2017, covering three seasons: spring (sampling time 1-4), summer (sampling time 5-10), and autumn (sampling time 11-14).
- Determination of environmental variables (water temperature, pH, conductivity, dissolved O<sub>2</sub>, total nitrogen, total phosphorus, total suspended solids, chlorophyl a, dissolved organic carbon) by field and laboratory measurements.
- Identification of abundant zooplankton species by microscopy.
- Determination of bacterial cell number by fluorescent microscopy.
- Filtration of water samples, DNA extraction, PCR amplification of 18S and 16S rRNA gene.
- Determination of the microeukaryotic and bacterial communities by Illumina amplicon sequencing.
- Bioinformatic analysis of the sequences with the mothur program.
- Data and statistical analysis using R.
- Networks were created using the Extended Local Similarity Analysis (eLSA) tool.
- Visualization of networks by the Cytoscape program.
- Identification of keystone OTUs by the weighted topological importance (WI) index.

## 3.2. Temporal dynamics of microbial diversity along environmental gradients

- Sampling of twenty-six soda pans of Seewinkel in two very contrasting spring season, 2017 (dry), 2018 (wet).
- Determination of environmental variables (water depth, Secchi depth, pH, conductivity, total nitrogen, total phosphorus, total suspended solids) by field and laboratory measurements.

- Filtration of water samples, DNA extraction, PCR amplification of 18S and 16S rRNA gene.
- Determination of the microeukaryotic and bacterial communities by Illumina amplicon sequencing.
- Bioinformatic analysis of the sequences with the mothur program.
- Creation of six larger groups: ciliates, fungi, eukaryotic phytoplankton, heterotrophic flagellates and nanoflagellates, cyanobacteria and bacteria (non-cyanobacteria).
- Data and statistical analysis using R.

## 4. Results and Discussion

### 4.1. Seasonal changes of the microeukaryotic and bacterial communities

Seasonality had a great impact on the microeukaryotic and bacterial communities of all pans. Various common seasonal dynamics of the microbial succession were observed, like the similarity of spring and autumn communities. Occasional droughts occurred during our sampling period, Kelemen-szék was dried out on five sampling occasions, constituting three distinct desiccation periods altogether, while Zab-szék had two distinct desiccation events. Every desiccation ended with refillment during the study. Desiccation and refillment combine physical and chemical stress on the microorganisms due to drastic changes in the habitat's water, salt, and nutrient content (Schimel, 2018). Based on our results, the microeukaryotic community was more sensitive to these changes, while the bacterial community remained more stable.

Overall, Bray-Curtis dissimilarities in community compositions between sampling times were greater for microeukaryotes on average, assuming the pans' core bacterial community has the ability to withstand sudden changes in environmental conditions. Core communities explained higher seasonal variance than noncore communities, which supported our hypothesis that seasonal adaptation is primarily driven through species recruitment from the core community.

Spring had similar properties in the five soda pans, although according to the community structure and interactions, it differed the most from summer and autumn. Spring was characterised by positive keystone OTUs (richly connected with positive associations to others). Positive keystone eOTUs abundant in spring belonged to flagellates like Colpodellida (Mylnikov, 2009) or parasitic taxa such as Cryptomycotina (Letcher et al., 2017) and Perkinsozoa (Mangot et al., 2011). Positive actinobacterial keystone OTUs were assigned to lineages like Luna1-A and acIV-C that have been identified in different aquatic habitats, including soda lakes (Ghai et al., 2012; Newton et al., 2011; Szabó et al., 2020).

After the analogous trajectory of spring, microbial community turnovers started to increase even for the turbid pans which were driven by the warming temperature of summer and shrinking habitat size. This was in agreement with previous studies showing that shrinking habitat size modifies community assembly and reduces stability (Bier et al., 2022). Desiccation periods were considered local stressors, due to the fact that desiccation is common in soda pans of this region, but not every pan dries out every year, and also different pans dry out in different years. Microeukaryotic communities turnover increased quite uniformly in the turbid pans and it increased extensively at each drying-rewetting cycle which suggested limited resilience against local stressors. It was reflected in the increased relative abundance of non-core4 after desiccation events, supporting the hypothesis that non-core OTUs are more important in the response to sudden environmental events.

All the networks had more positive associations than negative implying the dominance of positive interactions in the communities. Previous studies showed that positive correlations are more common in habitats characterized by high abiotic stress due to a higher number of mutualistic interactions letting species exist in harsher environments than otherwise would be possible (Hernandez et al., 2021; Travis et al., 2005). However, the dominance of positive associations (mutualisms) reduces network stability, especially in the case of low-modularity networks (Hernandez et al., 2021). Based on network topology, there was a clear difference between the colored Sós-ér, the two occasionally desiccated turbid pans (Kelemen-szék and Zab-szék), and the two turbid pans that did not dry out (Böddi-szék and Pan no. 60). The network of the soda pans reflected low community stability in these extreme habitats that was further aggravated by sudden environmental events (desiccation) and cyanobacteria blooms.

Overall, our results showed the studied soda pans despite drastic environmental changes and following community shifts are primarily populated by a common core microbiome and as a result of the identical climatic and meteorological conditions they have similar seasonal trends. However, the shared microbiome differed among pans of different habitat subtype (i.e., colored or turbid pan) and common seasonal trends were modified by local environmental events like desiccation. Adaptation to uniform seasonal changes was through species recruitment from the core community, while for microeukaryotes non-core members of the microbiome were involved in the response to sudden environmental events. Bacterial communities were resistant to stochastic events and adapted to the local stressors with species sorting from the core community and competitive exclusion.

### 4.2. Temporal dynamics of microbial diversity along environmental gradients

The dry spring of 2017 had more similar conditions to the reference summer data, while the wetter spring of 2018 was comparable to the reference spring data from a decade ago, indicating a seasonal shift in the aquatic environment. Based on our findings, conductivity was the most influential environmental variable shaping the microbial diversity patterns. Although the salinity gradient in the pans we studied was relatively narrow (0.6 to 11 mS/cm conductivity, equivalent to 0.4-8.8 g/L salinity, as per the conversion factor of (Boros et al., 2014)). Previous research suggested that significant changes in saline lake communities occur at lower salinity levels, particularly in the range of 3-10 g/L (Hammer, 1990; Schallenberg et al., 2003; Williams, 1987).

A previous review of microscopic data available from soda pans concluded that the number of eukaryotic phytoplankton species rarely exceeds 10 (Padisák and Naselli-Flores, 2021). In contrast, our study found an average eukaryotic phytoplankton OTU richness of 47 in 2017 and 49 in 2018, with a range of 16 (Runde Lacke in 2017) to 105 (Obere Höllacke in 2017). This

result shows that these habitats have a much higher diversity of phytoplankton than previously assumed, consistent with molecular results on the picocyanobacteria diversity of soda pans in Hungary and Austria (Felföldi et al., 2009; Somogyi et al., 2022). Only phytoplankton exhibited a U-shaped pattern in phylogenetic diversity across the salinity gradient in 2018, with the lowest diversity occurring at intermediate salinity (2.9 g/L), beyond which diversity increased again. Comparable U-shaped patterns of species richness in phytoplankton along salinity gradients have been observed from transitional gradients, ranging from freshwater to brackish to marine environments (Olli et al., 2022; 2019). However, in soda pans, the increase in phylogenetic diversity was not accompanied by a parallel increase in OTU richness, indicating that the pattern was most likely due to the disappearance of closely related taxa rather than the emergence of salt tolerant OTUs.

Our study found that HF-HNF richness and phylogenetic diversity increased exponentially with salinity, which is a pattern not previously reported from other saline systems, while previous research showed that heterotrophic flagellate diversity is primarily influenced by salinity and temperature (Azovsky et al., 2016; Je Lee and Patterson, 1998) and follows the "rule of critical salinity," meaning that species richness has a minimum in brackish waters (~5-8 g/L of salinity) (Tikhonenkov et al., 2006).

In conclusion, environmental variables had a stronger effect on community composition in the dry spring, and conductivity, TSS, and TP were the most important environmental variables affecting the diversity and community patterns across six major groups of microbes. In addition, the response of prokaryotes (Bacteria and Cyanobacteria) to the environmental gradients generally differed from the microeukaryotic groups.

## 5. Thesis

I.) How similar are the seasonal changes of planktonic microeukaryotic and prokaryotic communities in nearby soda pans?

The similarities of planktonic microbial communities from spring to autumn are altered by local effects (like occasional desiccation periods), which drives the community composition and structure by shifting the dominance and relationships of taxa. Therefore, the similar communities in the beginning of the year (spring) become more distinct when local effects are more pivotal (during summer and autumn).

II.) How core and non-core microbial taxa contribute to the adaptation of the microbial communities, and how does this contribution vary between microeukaryotic and bacterial communities?

Adaptation to seasonal changes differed between bacterial and microeukaryotic communities. For microeukaryotic communities non-core members of the microbiome were involved in the response to sudden environmental events, while bacterial communities adapted to extreme conditions through species sorting from the core community. III.) Does the identity and strength of the main environmental drivers change between subsequent years?

The environmental variables had a stronger effect on microbial community composition in the dry spring, than in the wet spring. The identity of the main environmental drivers remained the same in the two spring seasons and conductivity had the strongest effect in both years.

#### 6. References

- Azovsky, A.I., Tikhonenkov, D.V., Mazei, Y.A., 2016. An Estimation of the Global Diversity and Distribution of the Smallest Eukaryotes: Biogeography of Marine Benthic Heterotrophic Flagellates. Protist 167, 411–424. https://doi.org/10.1016/j.protis.2016.07.001
- Bier, R.L., Vass, M., Székely, A.J., Langenheder, S., 2022. Ecosystem size-induced environmental fluctuations affect the temporal dynamics of community assembly mechanisms. ISME J. 16, 2635–2643. https://doi.org/10.1038/s41396-022-01286-9
- Boros, E., Ecsedi, Z., Oláh, J., Szegedi, R., Dunn, J., 2013. Ecology and management of soda pans in the Carpathian Basin [WWW Document]. Hortobágy Environ. Assoc. URL https://mmebolt.hu/ecology-and-management-of-soda-pans-in-the-carpathian-basin-8207 (accessed 11.30.22).
- Boros, E., Horváth, Z., Wolfram, G., Vörös, L., 2014. Salinity and ionic composition of the shallow astatic soda pans in the Carpathian Basin. Ann. Limnol. 50, 59–69. https://doi.org/10.1051/limn/2013068
- Boros, E., V.-Balogh, K., Vörös, L., Horváth, Z., 2017. Multiple extreme environmental conditions of intermittent soda pans in the Carpathian Basin (Central Europe). Limnologica 62, 38–46. https://doi.org/10.1016/j.limno.2016.10.003
- Felföldi, T., 2020. Microbial communities of soda lakes and pans in the Carpathian Basin: a review. Biol. Futura 71, 393–404. https://doi.org/10.1007/s42977-020-00034-4
- Felföldi, T., Somogyi, B., Márialigeti, K., Vörös, L., 2009. Characterization of photoautotrophic picoplankton assemblages in turbid, alkaline lakes of the Carpathian Basin (Central Europe). J. Limnol. 68, 385. https://doi.org/10.4081/jlimnol.2009.385
- Ghai, R., Hernandez, C.M., Picazo, A., Mizuno, C.M., Ininbergs, K., Díez, B., Valas, R.,
  Dupont, C.L., McMahon, K.D., Camacho, A., Rodriguez-Valera, F., 2012.
  Metagenomes of mediterranean coastal lagoons. Sci. Rep. 2.
  https://doi.org/10.1038/srep00490
- Hammer, U.T., 1990. The effects of climate change on the salinity, water levels and biota of Canadian prairie saline lakes. SIL Proc. 1922-2010 24, 321–326. https://doi.org/10.1080/03680770.1989.11898751
- Hernandez, D.J., David, A.S., Menges, E.S., Searcy, C.A., Afkhami, M.E., 2021. Environmental stress destabilizes microbial networks. ISME J. 15, 1722–1734. https://doi.org/10.1038/s41396-020-00882-x

- Horváth, Z., Ptacnik, R., Vad, C.F., Chase, J.M., 2019. Habitat loss over six decades accelerates regional and local biodiversity loss via changing landscape connectance. Ecol. Lett. 22, 1019–1027. https://doi.org/10.1111/ele.13260
- Je Lee, W., Patterson, D.J., 1998. Diversity and Geographic Distribution of Free-Living Heterotrophic Flagellates – Analysis by PRIMER. Protist 149, 229–244. https://doi.org/10.1016/S1434-4610(98)70031-8
- Letcher, Peter M, Longcore, J.E., James, T.Y., Leite, D.S., Simmons, D.R., Powell, M.J., Letcher, P M, 2017. Morphology, Ultrastructure, and Molecular Phylogeny of Rozella multimorpha, a New Species in Cryptomycota. https://doi.org/10.1111/jeu.12452-4996
- Mangot, J.F., Debroas, D., Domaizon, I., 2011. Perkinsozoa, a well-known marine protozoan flagellate parasite group, newly identified in lacustrine systems: A review. Hydrobiologia 659, 37–48. https://doi.org/10.1007/s10750-010-0268-x
- Mylnikov, A.P., 2009. Ultrastructure and phylogeny of colpodellids (Colpodellida, Alveolata). Biol. Bull. 36, 582–590. https://doi.org/10.1134/S1062359009060065
- Newton, R.J., Jones, S.E., Eiler, A., McMahon, K.D., Bertilsson, S., 2011. A Guide to the Natural History of Freshwater Lake Bacteria. Microbiol. Mol. Biol. Rev. https://doi.org/10.1128/mmbr.00028-10
- Olli, K., Ptacnik, R., Klais, R., Tamminen, T., 2019. Phytoplankton Species Richness along Coastal and Estuarine Salinity Continua. Am. Nat. 194, E41–E51. https://doi.org/10.1086/703657
- Olli, K., Tamminen, T., Ptacnik, R., 2022. Predictable shifts in diversity and ecosystem function in phytoplankton communities along coastal salinity continua. Limnol. Oceanogr. Lett. lol2.10242. https://doi.org/10.1002/lol2.10242
- Padisák, J., Naselli-Flores, L., 2021. Phytoplankton in extreme environments: importance and consequences of habitat permanency. Hydrobiologia 848, 157–176. https://doi.org/10.1007/s10750-020-04353-4
- Pálffy, K., Felföldi, T., Mentes, A., Horváth, H., Márialigeti, K., Boros, E., Vörös, L., Somogyi, B., 2014. Unique picoeukaryotic algal community under multiple environmental stress conditions in a shallow, alkaline pan. Extremophiles 18, 111–119. https://doi.org/10.1007/s00792-013-0602-0
- Schallenberg, M., Hall, C., Burns, C., 2003. Consequences of climate-induced salinity increases on zooplankton abundance and diversity in coastal lakes. Mar. Ecol. Prog. Ser. 251, 181–189. https://doi.org/10.3354/meps251181
- Schimel, J.P., 2018. Life in Dry Soils: Effects of Drought on Soil Microbial Communities and Processes. Annu. Rev. Ecol. Evol. Syst. Annu Rev Ecol Evol Syst 12, 409–432. https://doi.org/10.1146/annurev-ecolsys-110617
- Somogyi, B., Felföldi, T., Boros, E., Szabó, A., Vörös, L., 2022. Where the Little Ones Play the Main Role—Picophytoplankton Predominance in the Soda and Hypersaline Lakes of the Carpathian Basin. Microorganisms 10, 818. https://doi.org/10.3390/microorganisms10040818
- Somogyi, B., Felföldi, T., Vanyovszki, J., Ágyi, Á., Márialigeti, K., Vörös, L., 2009. Winter bloom of picoeukaryotes in Hungarian shallow turbid soda pans and the role of light

and temperature. Aquat. Ecol. 43, 735–744. https://doi.org/10.1007/s10452-009-9269-0

- Sorokin, D.Y., Berben, T., Melton, E.D., Overmars, L., Vavourakis, C.D., Muyzer, G., 2014. Microbial diversity and biogeochemical cycling in soda lakes. Extremophiles 18, 791– 809. https://doi.org/10.1007/s00792-014-0670-9
- Szabó, A., Korponai, K., Somogyi, B., Vajna, B., Vörös, L., Horváth, Z., Boros, E., Szabó-Tugyi, N., Márialigeti, K., Felföldi, T., 2020. Grazing pressure-induced shift in planktonic bacterial communities with the dominance of acIII-A1 actinobacterial lineage in soda pans. Sci. Rep. 10, 1–13. https://doi.org/10.1038/s41598-020-76822-8
- Szabó, B., Szabó, A., Vad, C.F., Boros, E., Lukić, D., Ptacnik, R., Márton, Z., Horváth, Z., 2022.
  Microbial stowaways: Waterbirds as dispersal vectors of aquatic pro- and microeukaryotic communities. J. Biogeogr. 49, 1286–1298. https://doi.org/10.1111/jbi.14381
- Tikhonenkov, D.V., Mazei, Y.A., Mylnikov, A.P., 2006. Species diversity of heterotrophic flagellates in White Sea littoral sites. Eur. J. Protistol. 42, 191–200. https://doi.org/10.1016/j.ejop.2006.05.001
- Travis, J.M.J., Brooker, R.W., Dytham, C., 2005. The interplay of positive and negative species interactions across an environmental gradient: Insights from an individual-based simulation model. Biol. Lett. 1, 5–8. https://doi.org/10.1098/rsbl.2004.0236
- Williams, D.D., 1987. The Ecology of Temporary Waters. Springer Netherlands, Dordrecht. https://doi.org/10.1007/978-94-011-6084-1

### 7. Articles related to the dissertation

**Zsuzsanna Márton**, Bianka Csitári, Tamás Felföldi, Ferenc Jordán, András Hidas, Attila Szabó, Anna J Székely. (2023) Contrasting Response of Microeukaryotic and Bacterial Communities to the Interplay of Seasonality and Stochastic Events in Shallow Soda Lakes. - FEMS Microbiology Ecology, Accepted for publication

**Zsuzsanna Márton**, Beáta Szabó, Csaba F. Vad, Károly Pálffy, Zsófia Horváth. (2023) Environmental changes associated with drying climate are expected to affect functional groups of pro- and microuekaryotes differently in temporary saline waters. - Scientific Reports, 13:3243, https://doi.org/10.1038/s41598-023-30385-6

Beáta Szabó, Attila Szabó, Csaba F Vad, Emil Boros, Dunja Lukić, Robert Ptacnik, **Zsuzsanna Márton**, Zsófia Horváth. (2022) Microbial stowaways: Waterbirds as dispersal vectors of aquatic pro-and microeukaryotic communities. - Journal of Biogeography, 49:1286-1298, https://doi.org/10.1111/jbi.14381

## 8. Other papers

Kristóf Korponai, Sára Szuróczki, **Zsuzsanna Márton**, Attila Szabó, Paula V. Morais, Diogo Neves Proença, Erika Tóth, Emil Boros, Károly Márialigeti, Tamas Felfoldi. (2023) Habitat distribution of the Belliella genus in continental waters and the description of Belliella alkalica sp. nov., Belliella calami sp. nov. and Belliella filtrata sp. nov. - International Journal of Systematic and Evolutionary Microbiology, 73., https://doi.org/10.1099/ijsem.0.005928

Barna Putnoky-Csicsó, Szende Tonk, Attila Szabó, **Zsuzsanna Márton**, Franciska Tóthné Bogdányi, Ferenc Tóth, Éva Abod, János Bálint, Adalbert Balog. (2020) Effectiveness of the Entomopathogenic Fungal Species *Metarhizium anisopliae* Strain NCAIM 362 Treatments agains Soil Inhabiting *Melolontha melolontha* Larvae in Sweet Potato (*Ipomoea batatas* L.) - Journal of Fungi, 6:116, https://doi.org/10.3390/jof6030116.

Tamás Felföldi, **Zsuzsanna Márton**, Attila Szabó, Anikó Mentes, Károly Bóka, Károly Márialigeti, István Máthé, Mihály Koncz, Peter Schumann, Erika Tóth. (2019) *Siculibacillus lacustris* gen. nov., sp. nov., a new rosette-forming bacterium isolated from a freshwater crater lake (Lake St. Ana, Romania). - International Journal of Systematic and Evolutionary Microbiology, 69: 1731-1736, https://doi.org/10.1099/ijsem.0.003385